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(21) International Application Number: PCT/US92/01200 (22) International Filing Date: 13 February 1992 (13.02.92) (30) Priority data: 655,965 14 February 1991 (14.02.91) US (71) Applicant: THE GENERAL HOSPITAL CORPORATION [US/US]; Office of Technology Affairs, Thirteenth Street, Building 149, Suite 1101, Charlestown, MA 02129 (US). (72) Inventor: PODOLSKY, Daniel, K. ; 67 Yarmouth Road, Wellesley Hills, MA 02181 (US). (74) Agent: CLARK, Paul, T.; Fish & Richardson, 225 Franklin Street, Boston, MA 02110-2804 (US).		(81) Designated States: AT (European patent), AU, BE (European patent), CA, CH (European patent), DE (European patent), DK (European patent), ES (European patent), FR (European patent), GB (European patent), GR (European patent), IT (European patent), JP, LU (European patent), MC (European patent), NL (European patent), SE (European patent). Published <i>With international search report.</i>
(54) Title: INTESTINAL TREFOIL PROTEINS (57) Abstract Intestinal trefoil factors and nucleic acids encoding intestinal trefoil factors are disclosed. The intestinal trefoil factors disclosed are resistant to destruction in the digestive tract and can be used for the treatment of peptic ulcer diseases, inflammatory bowel diseases and other insults.		

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INTESTINAL TREFOIL PROTEINS

Background

The field of the invention is peptides useful for
5 treatment of disorders of the digestive system.

Jørgensen et al. (Regulatory Peptides 3:231, 1982)
describe a porcine pancreatic peptide, pancreatic
spasmolytic peptide (PSP). PSP was found to inhibit
"gastrointestinal motility and gastric acid secretion in
10 laboratory animal after parenteral as well as oral
administration." It was suggested that "if the results in
animal experiments can be confirmed in man, PSP may
posses a potential utility in treatment of gastroduodenal
ulcer diseases.

15 Summary of the Invention

In a first aspect, the invention features a
purified nucleic acid encoding an intestinal trefoil
factor (ITF).

In preferred embodiments, the intestinal trefoil
20 factor is mammalian intestinal trefoil factor, preferably
human, rat, bovine, or porcine intestinal trefoil factor.
In another preferred embodiment, the purified nucleic
acid encoding an intestinal trefoil factor is present
within a vector.

25 In a related aspect, the invention features a cell
that includes a vector encoding an intestinal trefoil
factor.

In another related aspect, the invention features
a substantially pure intestinal trefoil factor. In a
30 preferred embodiment, the polypeptide is detectably
labelled. In a related aspect, the invention features a
therapeutic composition that includes an intestinal
trefoil factor and a pharmacologically acceptable
carrier.

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In another aspect, the invention features a monoclonal antibody which preferentially binds (i.e., forms an immune complex with) an intestinal trefoil factor. In a preferred embodiment, the monoclonal antibody is detectably labelled.

In a related aspect, the invention features a method for detecting human intestinal trefoil factor in a human patient. The method includes the steps of contacting a biological sample obtained from the patient with a monoclonal antibody which preferentially binds intestinal trefoil factor, and detecting immune complexes formed with the monoclonal antibody. In preferred embodiments the biological sample is an intestinal mucosal scraping, or serum.

In a related aspect, the invention features a method for treating digestive disorders in a human patient, which method involves administering to the patient a therapeutic composition that includes an intestinal trefoil factor and a pharmacologically acceptable carrier.

In another aspect, the invention features a method for detecting binding sites for intestinal trefoil factor in a patient. The method involves contacting a biological sample obtained from the patient with the factor, and detecting the factor bound to the biological sample as an indication of the presence of the binding sites in the sample. By "binding sites", as used herein, is meant any antibody or receptor that binds to an intestinal trefoil factor protein, factor, or analog. The detection or quantitation of binding sites may be useful in reflecting abnormalities of the gastrointestinal tract.

In another aspect, the invention features substantially pure trefoil factor. In preferred

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embodiments, the intestinal trefoil factor is human, porcine, or bovine trefoil factor.

The term "intestinal trefoil factor" ("ITF") includes any protein which is substantially homologous to rat intestinal trefoil factor (Fig. 2, SEQ ID NO 2) and which is expressed in the large intestine, small intestine, or colon to a greater extent than it is expressed in tissues other than the small intestine, large intestine, or colon. Also included are: allelic variations; natural mutants; induced mutants; proteins encoded by DNA that hybridizes under high or low stringency conditions to ITF encoding nucleic acids retrieved from naturally occurring material; and polypeptides or proteins retrieved by antisera to ITF, especially by antisera to the active site or binding domain of ITF. The term also includes other chimeric polypeptides that include an ITF.

The term ITF also includes analogs of naturally occurring ITF polypeptides. Analogs can differ from naturally occurring ITF by amino acid sequence differences or by modifications that do not affect sequence, or by both. Analogs of the invention will generally exhibit at least 70%, more preferably 80%, more preferably 90%, and most preferably 95% or even 99%, homology with all or part of a naturally occurring ITF sequence. The length of comparison sequences will generally be at least about 8 amino acid residues, usually at least 20 amino acid residues, more usually at least 24 amino acid residues, typically at least 28 amino acid residues, and preferably more than 35 amino acid residues. Modifications include *in vivo*, or *in vitro* chemical derivatization of polypeptides, e.g., acetylation, or carboxylation. Also included are modifications of glycosylation, e.g., those made by modifying the glycosylation patterns of a polypeptide

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during its synthesis and processing or in further processing steps, e.g., by exposing the polypeptide to enzymes that affect glycosylation derived from cells that normally provide such processing, e.g., mammalian glycosylation enzymes. Also embraced are versions of the same primary amino acid sequence that have phosphorylated amino acid residues, e.g., phosphotyrosine, phosphoserine, or phosphothreonine. Analogs can differ from naturally occurring ITF by alterations of their primary sequence. These include genetic variants, both natural and induced. Induced mutants may be derived by various techniques, including random mutagenesis of the encoding nucleic acids using irradiation or exposure to ethanemethylsulfate (EMS), or may incorporate changes produced by site-specific mutagenesis or other techniques of molecular biology. See, Sambrook, Fritsch and Maniatis (1989), Molecular Cloning: A Laboratory Manual (2d ed.), CSH Press, hereby incorporated by reference. Also included are analogs that include residues other than naturally occurring L-amino acids, e.g., D-amino acids or non-naturally occurring or synthetic amino acids, e.g., β or γ amino acids.

In addition to substantially full-length polypeptides, the term ITF, as used herein, includes biologically active fragments of the polypeptides. As used herein, the term "fragment", as applied to a polypeptide, will ordinarily be at least about 10 contiguous amino acids, typically at least about 20 contiguous amino acids, more typically at least about 30 contiguous amino acids, usually at least about 40 contiguous amino acids, preferably at least about 50 contiguous amino acids, and most preferably at least about 60 to 80 or more contiguous amino acids in length. Fragments of ITF can be generated by methods known to those skilled in the art. The ability of a candidate

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fragment to exhibit a biological activity of ITF can be assessed by methods known to those skilled in the art. Also included in the term are biologically active ITF polypeptides containing amino acids that are normally removed during protein processing, including additional amino acids that are not required for the biological activity of the polypeptide, or including additional amino acids that result from alternative mRNA splicing or alternative protein processing events.

10 An ITF polypeptide, fragment, or analog is biologically active if it exhibits a biological activity of a naturally occurring ITF, e.g., the ability to alter gastrointestinal motility in a mammal.

 The invention also includes nucleic acid sequences, and purified preparations thereof, that encode the ITF polypeptides described herein. The invention also includes antibodies, preferably monoclonal antibodies, that bind specifically to ITF polypeptides.

 As used herein, the term "substantially pure" describes a compound, e.g., a nucleic acid, a protein, or a polypeptide, e.g., an ITF protein or polypeptide, that is substantially free from the components that naturally accompany it. Typically, a compound is substantially pure when at least 60%, more preferably at least 75%, more preferably at least 90%, and most preferably at least 99%, of the total material (by volume, by wet or dry weight, or by mole per cent or mole fraction) in a sample is the compound of interest. Purity can be measured by any appropriate method, e.g., in the case of polypeptides by column chromatography, polyacrylamide gel electrophoresis, or HPLC analysis.

 By "isolated DNA" is meant that the given DNA is free of the genes which, in the naturally-occurring genome of the organism from which the given DNA of the invention is derived, flank the given DNA. The term

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"isolated DNA" thus encompasses, for example, cDNA, cloned genomic DNA, and synthetic DNA. A "purified nucleic acid", as used herein, refers to a nucleic acid sequence that is substantially free of other

5 macromolecules (e.g., other nucleic acids and proteins) with which it naturally occurs within a cell. In preferred embodiments, less than 40% (and more preferably less than 25%) of the purified nucleic acid preparation consists of such other macromolecules.

10 "Homologous", as used herein, refers to the subunit sequence similarity between two polymeric molecules, e.g., between two nucleic acid molecules, e.g., two DNA molecules, or two polypeptide molecules. When a subunit position in both of the two molecules is

15 occupied by the same monomeric subunit, e.g., if a position in each of two DNA molecules is occupied by adenine, then they are homologous at that position. The homology between two sequences is a direct function of the number of matching or homologous positions, e.g., if

20 half, e.g., 5 of 10, of the positions in two compound sequences are homologous then the two sequences are 50% homologous, if 90% of the positions, e.g., 9 of 10, are matched or homologous the two sequences share 90% homology. By way of example, the DNA sequences

25 3'ATTGCC'5 and 3'TATGGC'5 share 50% homology. By "substantially homologous" is meant largely but not wholly homologous.

The ITF proteins of the invention are resistant to destruction in the digestive tract, and can be used for

30 treatment of peptic ulcer diseases, inflammatory bowel diseases, and for protection of the intestinal tract from injury caused by bacterial infection, radiation injury or other insults. An ITF protein, fragment, or analog can also be used to treat neoplastic cancer.

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Other features and advantages of the invention will be apparent from the following description of the preferred embodiments thereof, and from the claims.

Detailed Description

5 The drawings will first be briefly described.

Drawings

Figure 1 is a depiction of the nucleotide sequence of rat trefoil factor (SEQ ID NO: 1).

10 Figure 2 is a depiction of the deduced amino acid sequence of rat trefoil factor (SEQ ID NO: 2).

Figure 3 is a depiction of the amino acid sequences of rat trefoil factor, pS2 protein, and pancreatic spasmolytic polypeptide. The sequences are aligned so as to illustrate the amino acid sequence
15 homology between the proteins. Dashes (-) indicate the insertion of spaces which improve alignment. Bars () indicate sequence identities.

Figure 4 depicts the disulfide bond structure proposed for pS2 (panel A) and PSP (panel B);

20 Figure 5 is a depiction of the proposed disulfide bond structure of rat intestinal trefoil factor.

Figure 6 is a depiction of the nucleotide sequence of the human intestinal trefoil factor cDNA and the corresponding deduced amino acid sequence (SEQ ID NO: 3).

25 Purification and cloning of rITF

An inhibitor of soft agar colony formation by human breast carcinoma-derived BT-20 cells (ATTC HTB79) was isolated from cytology-positive human malignant effusions (Podolsky et al., *Cancer Res.* 48:418, 1988;
30 hereby incorporated by reference). The factor also inhibited soft agar colony formation by human colon carcinoma-derived HCT15 cells (ATTC-CCL225). Inhibition was not observed for polyoma and murine sarcoma virus transformed rodent fibroblast lines. The isolated factor
35 (transformed cell-growth inhibiting factor or TGIF) had

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an apparent molecular weight of 110,000 kD and appeared to consist of two 55,000 kD subunits linked by sulphhydryl bonds.

The purified protein was partially sequenced. The
5 sequence from the amino terminal 14 amino acids was used to produce a set of degenerate oligonucleotide probes for screening of a rat intestinal epithelial cell cDNA library.

A rat intestinal cDNA library (Lambda ZAP[®] II,
10 Stratagene, La Jolla, CA) was produced by standard techniques (Ausubel et al., eds., *Current Protocols in Molecular Biology*, John Wiley & Sons, New York, 1989) using cells purified by the method of Weisner (*J. Biol Chem.* 248:2536, 1973). Screening of the cDNA library with
15 the fully degenerate oligonucleotide probe described above resulted in the selection of 21 clones. One of the clones (T3411) included a core sequence which encoded a single open reading frame. The nucleotide sequence of the open reading frame and flanking DNA is presented in Fig.
20 1 (SEQ ID NO 1). The insert present in T3411 was nick translated (Ausubel et al., *supra*) to produce a radioactively labelled probe for Northern blot analysis of rat poly(A)⁺ RNA. Northern analysis demonstrated that
25 RNA corresponding to the cloned cDNA fragment was expressed in small intestine, large intestine, and kidney; no expression was detected in the lung, spleen, heart, testes, muscle, stomach, pancreas, or liver. In the tissues in which the RNA was expressed, the level was comparable to that of actin.

30 The open reading frame of clone T3411 encoded an 81 amino acid peptide (Fig. 2; SEQ ID NO 2). Comparison of the sequence of the encoded peptide, referred to as rat intestinal trefoil factor (rITF), to the sequence of proteins in the Gen bank database revealed significant
35 homology to human breast cancer associated peptide (pS2;

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Jakowlev et al., *Nucleic Acids Res.* 12:2861, 1984) and porcine pancreatic spasmolytic peptide (PSP; Thim et al., *Biochem. Biophys. Acta* 827:410, 1985). Fig. 3 illustrates the homology between rITF, PSP and pS2. Porcine

5 pancreatic spasmolytic factor (PSP) and pS2 are both thought to fold into a characteristic structure referred to as a trefoil. A trefoil structure consists of three loops formed by three disulfide bonds. pS2 is thought to include one trefoil (Fig. 4A), and PSP is thought to

10 include two trefoils (Fig. 4B). The region of rITF (nucleotide 114 to nucleotide 230 which encodes cys to phe) which is most similar to PSP and pS2 includes six cysteines all of which are in the same position as the cysteines which make up the trefoil in pS2 (Fig. 3). Five

15 of these six cysteines are in the same position as the cysteines which form the amino terminal trefoil of PSP (Fig. 3). Fig. 5 depicts the proposed disulfide bond configuration of rITF.

Based on homology to PSP and pS2 (Mori et al., *Biochem. Biophys. Res. Comm.* 155:366, 1988; Jakowlev et al., *Nucleic Acids Res.* 12:2861, 1984), rITF includes a presumptive pro- sequence (met¹ to ala²²) in which 12 of 22 amino acids have hydrophobic side chains.

Production of Anti-rITF Antibodies

25 A peptide corresponding to the carboxy-terminal 21 amino acids of rITF was synthesized and coupled to bovine serum albumin (BSA). This conjugate (and the unconjugated peptide) was used to raise polyclonal antibodies in rabbits. All procedures were standard protocols such as

30 those described in Ausubel et al. (*supra*). The anti-rITF antibodies were used in an indirect immunofluorescence assay for visualization of rITF in rat tissues. Cryosections of rat tissues were prepared using standard techniques, and fluorescein labelled goat anti-rabbit monoclonal antibody

35 (labelled antibodies are available from such suppliers

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Kirk gaard and Perry Laboratories, Gaithersberg, MD; and Bioproducts for Science, In., Indianapolis, IN) was used to detect binding of rabbit anti-rITF antibodies. By this analysis rITF appears to be present in the globlet cells of the small intestine but not in the stomach or the pancreas.

Cloning of Human Intestinal Trefoil Factor

DNA encoding the rat intestinal trefoil factor can be used to identify a cDNA clone encoding the human intestinal trefoil factor (hITF). This can be accomplished by screening a human colon cDNA library with a probe derived from rITF or with a probe derived from part of the hITF gene. The latter probe can be obtained from a human colon or intestinal cDNA using the polymerase chain reaction to isolate a part of the hITF gene. This probe can then serve as a specific probe for the identification of clones encoding all of the hITF gene.

Construction of a cDNA Library.

A human colon or intestinal cDNA library in λ gt10 or λ gt11, or some other suitable vector is useful for isolation of hITF. Such libraries may be purchased (Clontech Laboratories, Palo Alto, CA: HLI034a, HLI0346b). Alternatively, a library can be produced using mucosal scrapings from human colon or intestine. Briefly, total RNA is isolated from the tissue essentially as described by Chirgwin et al. (*Biochemistry* 18:5294, 1979; see also Ausubel et al., *supra*). An oligo (dT) column is then used to isolate poly(A)⁺ RNA by the method of Aviv et al. (*J. Mol. Biol.* 134:743, 1972; see also Ausubel et al., *supra*). Double-stranded cDNA is then produced by reverse transcription using oligo (dT)₁₂₋₁₈ or random hexamer primers (or both). RNase H and *E. coli* DNA pol I are then used to replace the RNA strand with a second DNA strand. In a subsequent step *E. coli* DNA ligase and T4

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DNA polymerase are used to close gaps in the second DNA strand and create blunt ends. Generally, the cDNA created is next methylated with EcoRI methylase and EcoRI linkers are added (other linkers can be used depending on the vector to be used). In subsequent steps the excess linkers are removed by restriction digestion and the cDNA fragments are inserted into the desired vector. See Ausubel et al., *supra* and Sambrook et al. (*Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1990) for detailed protocols. Useful vectors include: λ gt11, λ gt10, Lambda ZAP[®] II vector, Lambda Uni-ZAP[™] XR vector, all available from Stratagene (La Jolla, CA).

The cDNA library must be packaged into phage; this is most readily accomplished by use of a commercial *in vitro* packaging kit, e.g., Gigapack[®] II Gold or Gigapack[®] II Plus (Stratagene, La Jolla, CA). See Ausubel et al. (*supra*) for packaging protocols and suitable host strains. The library is preferably amplified soon after packaging; this step generates sufficient clones for multiple screening of the library. See Ausubel et al. *supra* or Sambrook et al. *supra* for details of amplification protocols and procedures for storing the amplified library.

Screening of the cDNA Library. To screen the library it must be placed on an appropriate host strain (e.g., Y1090 or Y1088 for λ gt10 libraries, C600hflA for λ gt10 libraries). After plating the phage, plaques are transferred to nitrocellulose or nylon filters (See Ausubel et al., *supra* and Sambrook et al. *supra*). The filters are then probed with $\alpha^{32}\text{P}$ -labelled nick translated probe derived from rITF. The probe is preferentially generated using a portion of the region of rITF DNA coding for the trefoil structure (nucleotides 114 to 230 of SEQ ID NO. 1 which encode cyS32 to ph⁷¹ of SEQ ID NO.

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2). This region is conserved between rITF, ps2 and PSP, and it is likely that this region is conserved between rITF and hITF. Once a plaque is identified several cycles of plaque purification are required to isolate a pure
5 clone encoding hITF. A phage DNA isolation is performed and the cDNA insert can be subcloned into an appropriate vector for restriction mapping and sequencing. If the phage vector is Lambda ZAP® II, coinfection with helper
10 SK⁻ phagemid vector (Stratagene, La Jolla, CA) harboring the cDNA; alternatively the phage clone is purified and the cDNA insert is subcloned into a vector suitable for restriction mapping and sequencing. If the clone does
15 not contain the entire hITF gene (as assessed by homology to rITF and the presence of start and stop codons), the library can be rescreened with the original rITF probe or, preferably, with a probe generated from the hITF clone obtained. If none of the clones contain the intact
20 overlapping fragments of hITF.

Direct Isolation of an hITF Probe by PCR

It is possible to isolate part of the hITF gene directly from the packaged library or cDNA. To isolate a portion of hITF directly from the packaged library, a
25 pair of oligonucleotide primers and Taq polymerase are used to amplify the DNA corresponding to the hITF gene. The primers used would be approximately 15-20 nucleotides long and correspond in sequence to the 5'-most and 3'-most portions of the rITF coding sequence. Friedman et
30 al. (in *PCR Protocols: A Guide to Methods and Applications*, Innis et al., eds., Academic Press, San Diego) describe a procedure for such amplification. Briefly, phage particles are disrupted by heating; Taq polymerase, primers (300 pmol of each), dNTPs, and Taq
35 polymerase buffer are added; and the mixture is thermally

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cycled to amplify DNA. The amplified DNA is isolated by agarose gel electrophoresis. The ends of the fragment are prepared for ligation into an appropriate vector by making them flush with T4 polymerase and, if desired, adding linkers. Alternatively, a restriction site may be engineered into the fragment by using primers which have sequence added to their 5' ends which sequence will generate an appropriate sticky end when digested. For example the sequence: 5'-GGGCGGCCGC3' can be added to the 5' end of each primer. This sequence includes the NotI restriction site flanked at the 5' end by the sequence: GG. The additional nucleotides prevent the 5' ends from denaturing and interfering with subsequent restriction digestion with NotI. The gel purified DNA of the appropriate size is next cloned into a cloning vector for sequencing and restriction mapping. This clone will not have the entire hITF sequence, rather it will be a combination of hITF (the region between the sequences corresponding to the primers) and rITF (the 5' and 3' ends which correspond to the primer sequences). However, this DNA can be used to generate a labelled probe (produced by nick translation or random primer labelling) which, since it is the correct hITF sequence, can be used in a high stringency screening of the library from which the cDNA was originally isolated. In an alternative approach, cDNA can be used in the above procedure instead of a packaged library. This eliminates the steps of modifying the cDNA for insertion into a vector as well as cDNA packaging and library amplification. Ausubel et al. supra provides a protocol for amplification of a particular DNA fragment directly from cDNA and a protocol for amplification from poly(A)⁺ RNA.

Identification of a Presumptive Human ITF clone

A nick translated probe derived from rITF cDNA (corresponding to nucleotides 1 to 431 of SEQ ID No. 1)

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was used for Northern blot analysis of poly(A)⁺ RNA derived from human intestinal mucosal scrapings. Probe hybridization and blot washing were carried out according to standard procedures. Probe (5×10^5 cpm/ml hybridization buffer) was hybridized to the filter at 45°C in 5X SSC with 30% formamide. The filter was then washed at 60°C in 5X SSC with 40% formamide. Using this protocol a band was clearly visible after an overnight exposure of the filter with an intensifying screen. This result indicated that there is sufficient homology between rITF and hITF to allow the use of probes derived from the sequence of the rITF gene for identification of the hITF gene.

A human intestinal cDNA library was obtained from Clontech (Palo Alto, CA). Alternatively, a human intestinal cDNA library may be produced from mucosal scrapings as described above. Four oligonucleotide probes were selected for screening the library cDNA. Two of the probes correspond to sequences within the region of rITF encoding the trefoil and are referred to as internal probes (5'-gtacattctgtctcttgacaga-3' and 5'-taaccctgctgctgctggtcctgg3'). The other two probes recognize sequences within rITF but outside of the trefoil encoding region and are referred to as external probes (5'-gtttgcgtgctgcatggaga-3' and 5'-ccgcaattagaacagccttgt-3'). These probes were tested for their utility by using them to screen the rat intestinal cDNA library described above. Each of the four probes could be used to identify a clone harboring all or part of the rITF gene. This result indicates that these probes may be used to screen the human intestinal library for the presence of hITF.

The internal probes were used as described above to amplify a DNA fragment from human colon library cDNA (Clontech, Palo Alto, CA). Linkers were added to the

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isolated DNA fragment which was then inserted into pBluescript phagemid vector (Stratagene, La Jolla, CA). The region of this clone corresponding to the sequence of human cDNA (i.e., not including the sequence

5 corresponding to the internal probes) was used to make a radioactively labelled probe by random oligonucleotide-primed synthesis (Ausebel et al., *supra*). This probe was then used to screen the human colon cDNA library. This screening led to the identification of 29 clones. One of
10 these clones (HuPCR-ITF) was nick-translated to generate a probe for Northern analysis of poly(A)⁺ RNA isolated from human intestinal mucosal scrapings. A single band of roughly the same size as the rat transcript (approximately 0.45 kD) was observed.

15 Northern analysis of poly(A)⁺ isolated from human tissues indicated that RNA corresponding to this probe was expressed in the small intestine and the large intestine but not in the stomach or the liver. These results indicate that the clone does not encode the human
20 homolog of porcine PSP. Porcine PSP is expressed in porcine pancreas and is not significantly expressed in the small or large intestine. These results also distinguish the cloned gene from ps2 which is expressed in the stomach.

25 Figure 6 shows the nucleic acid sequence information for human ITF cDNA, along with the deduced amino acid sequence in one-letter code (SEQ ID NO: 3). This clone was obtained by the methods described above.
Production of hITF

30 The isolated hITF gene can be cloned into a mammalian expression vector for protein expression. Appropriate vectors include pMAMneo (Clontech, Palo Alto, CA) which provides a RSV-LTR enhancer linked to a dexamethasone-inducible MMTV-LTR promoter, an SV40 origin
35 of replication (allows replication in COS cells), a

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neomycin gene, and SV40 splicing and polyadenylation sites. This vector can be used to express the protein in COS cells, CHO cells, or mouse fibroblasts. The gene may also be cloned into a vector for expression in drosophila cells using the bacoluvirus expression system.

Purification of Intestinal Trefoil Factor

Intestinal trefoil factor can be purified from intestinal mucosal scrapings of human, rats or any other species which expresses ITF (pigs and cows may provide a source of ITF). The purification procedure used for PSP will be useful for the purification of ITF since the proteins are likely to be homologous. Jorgensen et al. describes a method for purification of PSP (Regulatory Peptides 3:207, 1982). The preferred method is the second approach described by Jorgensen et al. (*supra*). This method involves chromatography of SP-Sephadex C-25 and QAE Sephadex A-25 columns (Sigma, St. Louis, MO) in acidic buffer. Anti-Intestinal Trefoil Factor Monoclonal Antibodies

Anti-intestinal trefoil factor monoclonal antibodies can be raised against synthetic peptides whose sequences are based on the deduced amino acid sequence of cloned hITF (SEQ ID NO: 3). Most commonly the peptide is based on the amino-or carboxy-terminal 10-20 amino acids of the protein of interest (here hITF). The peptide is usually chemically cross-linked to a carrier molecule such as bovine serum albumin or keyhole limpet hemocyanin. The peptide is selected with the goal of generating antibodies which will cross-react with the native hITF. Accordingly, the peptide should correspond to an antigenic region of the peptide of interest. This is accomplished by choosing a region of the protein which is (1) surface exposed, e.g., a hydrophobic region or (2) relatively flexible, e.g., a loop region or a β -turn region. In any case, if the peptide is to be coupled to

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a carrier, it must have an amino acid with a side chain capable of participating in the coupling reaction. See Hopp et al. (*Mol. Immunol.* 20:483, 1983; *J. Mol. Biol.* 157:105, 1982) for a discussion of the issues involved in the selection of antigenic peptides. A second consideration is the presence of a protein homologous to hITF in the animal to be immunized. If such a protein exists, it is important to select a region of hITF which is not highly homologous to that homolog.

10 For hITF, peptides that correspond to the amino-terminal or carboxy-terminal 15 amino acids are likely to be less homologous across species and exposed to the surface (and thus antigenic). Thus they are preferred for the production of monoclonal antibodies. Purified hITF
15 can also be used for the generation of antibodies.

Use

In the practice of the present invention ITF may be administered orally, intravenously, or intraperitoneally for treatment of peptic ulcer diseases, inflammatory bowel diseases, and for protection of the
20 intestinal tract from injury caused by bacterial infection, radiation injury or other insults. The mode of administration, dosage, and formulation of ITF depends upon the condition being treated.

25 Other Embodiments

Other embodiments are within the following claims. For example, ITF may be used to produce monoclonal antibodies for the detection of ITF in intestinal tissue or blood serum by means of an indirect immunoassay. ITF
30 may be detectably labelled and used in an *in situ* hybridization assay for the detection of ITF binding sites. Labels may include, but are not limited to, fluorescein or a radioactive ligand.

ITF may be used to protect and stabilize other
35 proteins. This protection is accomplished by forming a

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hybrid molecule in which all or part of ITF is fused to either the carboxy-terminus or the amino-terminus (or both) of the protein of interest. Because ITF is resistant to degradation in the digestive system, it will
5 protect the protein of interest from such degradation. As a consequence, the protein of interest is likely to remain active in the digestive system and/or will be more readily absorbed in an intact form.

What is claimed is:

SEQUENCE LISTING**(1) GENERAL INFORMATION:**

(i) APPLICANT: Podolsky, Daniel K.

(ii) TITLE OF INVENTION: INTESTINAL TREFOIL FACTORS

(iii) NUMBER OF SEQUENCES: 3

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(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: 3.5" Diskette, 1.44 Mb
(B) COMPUTER: IBM PS/2 Model 50Z or
55SX
(C) OPERATING SYSTEM: IBM P.C. DOS (Version
3.30)
(D) SOFTWARE: WordPerfect (Version 5.0)

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER:
(B) FILING DATE:
(C) CLASSIFICATION:

(vii) PRIOR APPLICATION DATA:

(A) APPLICATION NUMBER:
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(viii) ATTORNEY/AGENT INFORMATION:

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(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 1:

- 20 -

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 431
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

GAAGTTTGCG TGCTGCC 17

ATG GAG ACC AGA GCC TTC TGG ATA ACC CTG CTG CTG GTC CTG GTT GCT 65
 Met Glu Thr Arg Ala Phe Trp Ile Thr Leu Leu Leu Val Leu Val Ala
 1 5 10 15

GGG TCC TCC TGC AAA GCC CAG GAA TTT GTT GGC CTA TCT CCA AGC CAA 113
 Gly Ser Ser Cys Lys Ala Gln Glu Phe Val Gly Leu Ser Pro Ser Gln
 20 25 30

TGT ATG GCG CCA ACA AAT GTC AGG GTG GAC TGT AAC TAC CCC ACT GTC 161
 Cys Met Ala Pro Thr Asn Val Arg Val Asp Cys Asn Tyr Pro Thr Val
 35 40 45

ACA TCA GAG CAG TGT AAC AAC CGT GGT TGC TGT TTT GAC TCC AGC ATC 209
 Thr Ser Glu Gln Cys Asn Asn Arg Gly Cys Cys Phe Asp Ser Ser Ile
 50 55 60

CCA AAT GTG CCC TGG TGC TTC AAA CCT CTG CAA GAG ACA GAA TGT ACA 257
 Pro Asn Val Pro Trp Cys Phe Lys Pro Leu Gln Glu Thr Glu Cys Thr
 65 70 75 80

TTT 260
 Phe

TGAAGCTGTC CAGGCTCCAG GAAGGGAGCT CCACACCCTG GACTCTTGCT GATGGTAGTG 320

GCCCAGGGTA ACACTCACCC CTGATCTGCT CCCTCGCGCC GGCCAATATA GGAGCTGGGA 380

GTCCAGAAGA ATAAAGACCT TACAGTCAGC ACAAGGCTGT TCTAATTGCG G 431

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 81
 (B) TYPE: amino acid
 (C) STRANDEDNESS: N/A
 (D) TOPOLOGY: N/A

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

[illegible]

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER:

3: human

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 403
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

G

ATG	CTG	GGG	CTG	GTC	CTG	GCC	TTC	CTG	TCC	TCC	AGC	TCT	GCT	GAG	GAG
Met	Leu	Gly	Leu	Val	Leu	Ala	Leu	Leu	Ser	Ser	Ser	Ser	Ala	Glu	Glu
1				5					10					15	
TAC	GTG	GGC	CTG	TCT	GCA	AAC	CAG	TGT	GCC	GTG	CCG	GCC	AAG	GAC	AGG
Tyr	Val	Gly	Leu	Ser	Ala	Asn	Gln	Cys	Ala	Val	Pro	Ala	Lys	Asp	Arg
			20					25					30		
GTG	GAC	TGC	GGC	TAC	CCC	CAT	GTC	ACC	CCC	AAG	GAG	TGC	AAC	AAC	CGG
Val	Asp	Cys	Gly	Tyr	Pro	His	Val	Thr	Pro	Lys	Glu	Cys	Asn	Asn	Arg
		35					40					45			
GGC	TGC	TGC	TTT	GAC	TCC	AGG	ATC	CCT	GGA	GTG	CCT	TGG	TGT	TTC	AAG
Gly	Cys	Cys	Phe	Asp	Ser	Arg	Ile	Pro	Gly	Val	Pro	Trp	Cys	Phe	Lys
	50					55					60				

CCC CTG ACT AGG AAG ACA GAA TGC ACC TTC
Pro Leu Thr Arg Lys Thr Glu Cys Thr Phe
65 70

**TGAGGCACCT CCAGCTGCCC CTGGGATGCA GGCTGAGCAC CCTTGCCCGG CTGTGATTGC
 TGCCAGGCAC TGTTTCATCTC AGTTTTTCTG TCCCTTTGCT CCCGGCAAGC TTTCTGCTGA
 AAGTTCATAT CTGGAGCCTG ATGTCTTAAC GAATAAAGGT CCCATGCTCC ACCCGAAAAA**

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403

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Claims

- 1 1. A purified nucleic acid encoding an intestinal
2 trefoil factor.
- 1 2. The purified nucleic acid of claim 1 wherein said
2 intestinal trefoil factor is mammalian intestinal trefoil factor
- 1 3. The purified nucleic acid of claim 2 wherein said
2 mammal is a human.
- 1 4. The purified nucleic acid of claim 2 wherein said
2 mammal is a rat.
- 1 5. The purified nucleic acid of claim 2 wherein said
2 mammal comprises a cow, or a pig.
- 1 6. The purified nucleic acid of claim 1, said purified
2 nucleic acid being present within a vector.
- 1 7. A cell comprising a vector encoding an intestinal
2 trefoil factor.
- 1 8. A substantially pure intestinal trefoil factor.
- 1 9. A therapeutic composition comprising said factor of
2 claim 8 and a pharmacologically acceptable carrier.
- 1 10. A monoclonal antibody which preferentially binds the
2 factor of claim 8.
- 1 11. The monoclonal antibody of claim 10 wherein said
2 monoclonal antibody is detectably labelled.

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1 12. A method for detecting intestinal trefoil factor in
2 a human patient comprising
3 contacting a biological sample obtained from said patient
4 with said monoclonal antibody of claim 10, and
5 detecting immune complexes formed with said monoclonal
6 antibody.

1 13. The method of claim 12 wherein said biological
2 sample is an intestinal mucosal scraping.

1 14. The method of claim 12 wherein said biological
2 sample is serum.

1 15. A method for treating digestive disorders in a human
2 patient, comprising
3 administering to said patient the therapeutic
4 composition of claim 9.

1 16. The factor of claim 8 wherein said factor is
2 detectably labelled.

1 17. A method for detecting binding sites for intestinal
2 trefoil factor in a patient comprising
3 contacting a biological sample obtained from said pati nt
4 with said factor of claim 8, and
5 detecting said factor bound to said biological sample as
6 an indication of the presence of said binding sites in said
7 sample.

1 18. An isolated DNA comprising a sequence encoding an
2 intestinal trefoil factor.

1 19. The isolated DNA of claim 18, wherein said DNA is
2 present within a vector.

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1 20. The isolated DNA of claim 18, wherein said
2 intestinal trefoil factor is a mammalian intestinal trefoil
3 factor.

1 21. The isolated DNA of claim 20, wherein said mammal is
2 a human.

1 22. The isolated DNA of claim 20, wherein said mammal is
2 a rat.

1 23. The isolated DNA of claim 20, wherein said mammal
2 comprises a cow, or a pig.

FIG. 1

gaagtttgcg	tgctgcc	17
atg gag acc aga gcc ttc tgg ata acc ctg ctg gtc ctg gtt		32
gct ggg tcc tcc tgc aaa gcc cag gaa ttt gtt ggc cta tct cca		77
agc caa tgt atg gcg cca aca aat gtc agg gtg gac tgt aac tac		122
ccc act gtc aca tca gag cag tgt aac aac cgt ggt tgc tgt ttt		167
gac tcc agc atc cca aat gtg ccc tgg tgc ttc aaa cct ctg caa		212
gag aca gaa tgt aca ttt		230
tgaagctgtc caggctccag gaaggagct ccacaccctg gactcttgct		280
gatggtagtg gcccagggtg aactcaccct ctgatctgct ccctcgcgcc		330
ggccaatata ggagctggga gtccagaaga ataaagacct tacagtcagc		380
acaaggctgt tctaattgcg g		401

Met Glu Thr Arg Ala Phe Trp Ile Thr Leu Leu Leu Val Leu Val
 5 10 15
 Ala Gly Ser Ser Cys Lys Ala Gln Glu Phe Val Gly Leu Ser Pro
 20 25 30
 Ser Gln Cys Met Ala Pro Thr Asn Val Arg Val Asp Cys Asn Tyr
 35 40 45
 Pro Thr Val Thr Ser Gly Gln Cys Asn Asn Arg Gly Cys Cys Phe
 50 55 60
 Asp Ser Ser Ile Pro Asn Val Pro Trp Cys Phe Lys Pro Leu Gln
 65 70 75
 Glu Thr Glu Cys Thr Phe
 80

FIG. 2

rITF METRAFWITLLVLVAGSSCKAQEFVGLSPSQCMAPTNNVRVDCNYPTVTSEQCNRGCCC
 pS2 -----EAQ-----TETCTVAPRERQNCGFPGVTPSQCANRKGCC
 PSP -----EKPAACRCSRQDPKN-RVNCGFPGITSDQCFTSGCC
 rITF FDSSIPNVPWCFK-----PLQ-----ETECT-----F
 pS2 FDDTVRGVPWCFY-----PNTIDVPPEECE-----F
 PSP FDSQVPGVPWCFK-----PLP-----AQSEECVMEV

FIG. 3

FIG. 4a

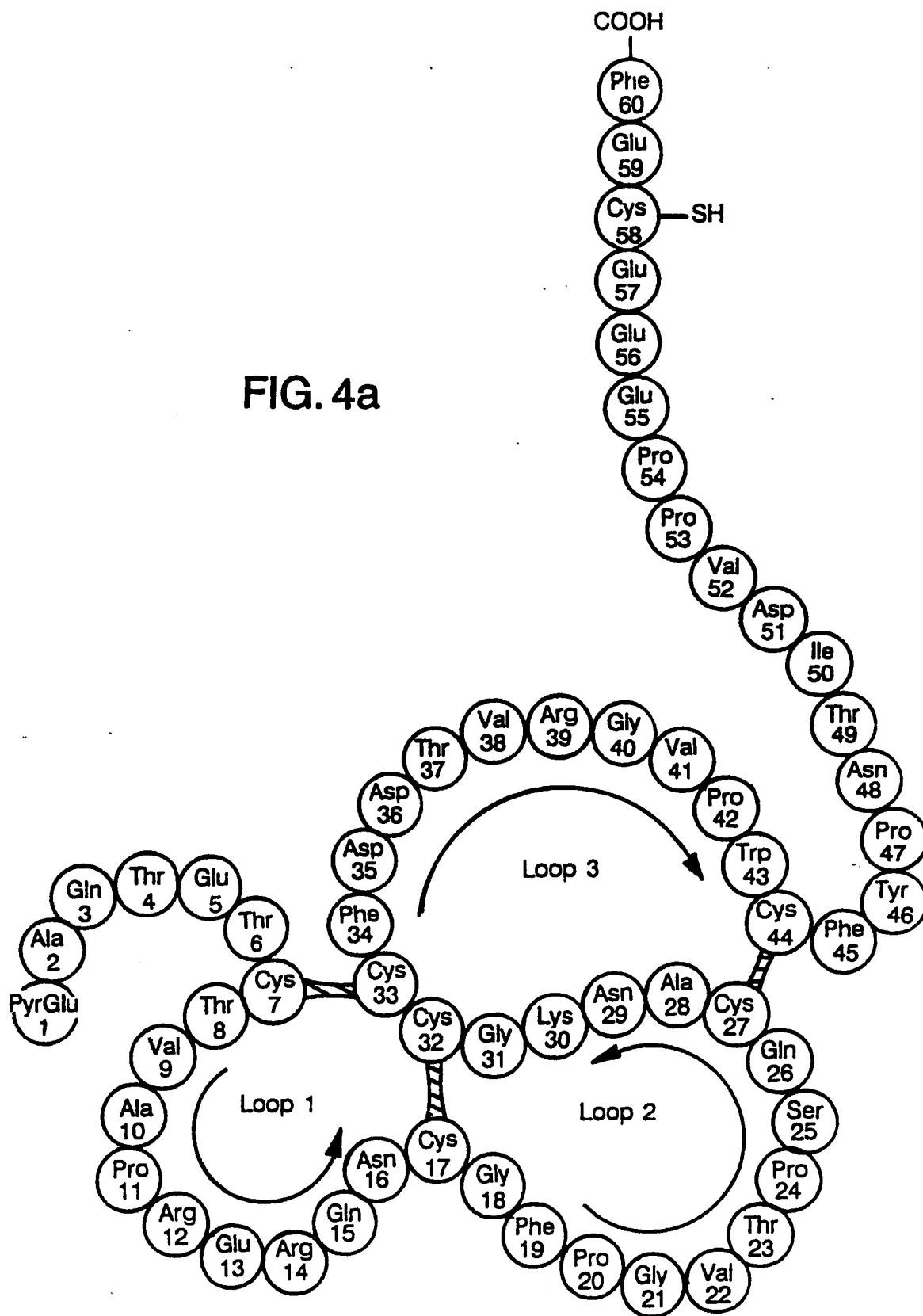
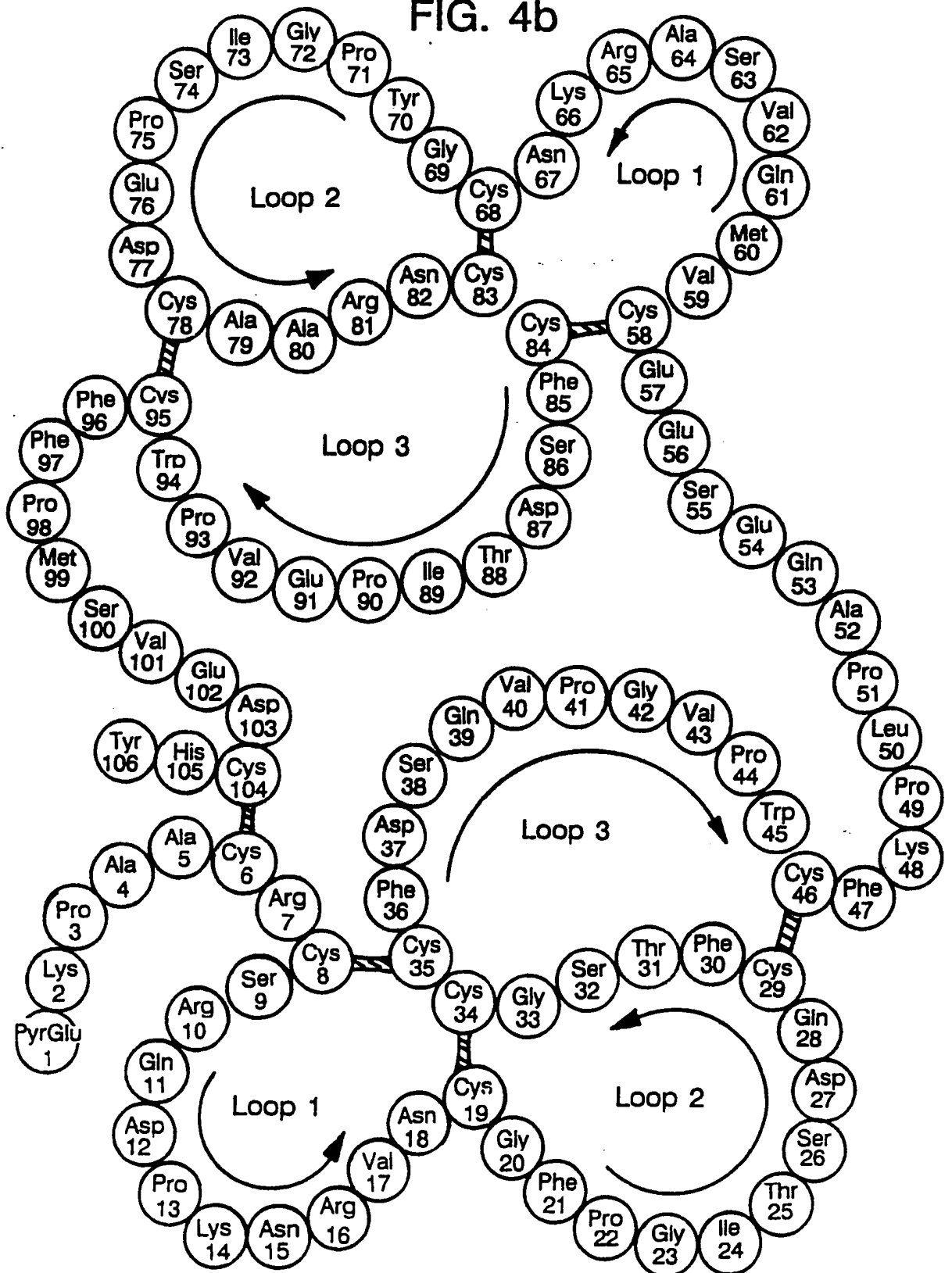
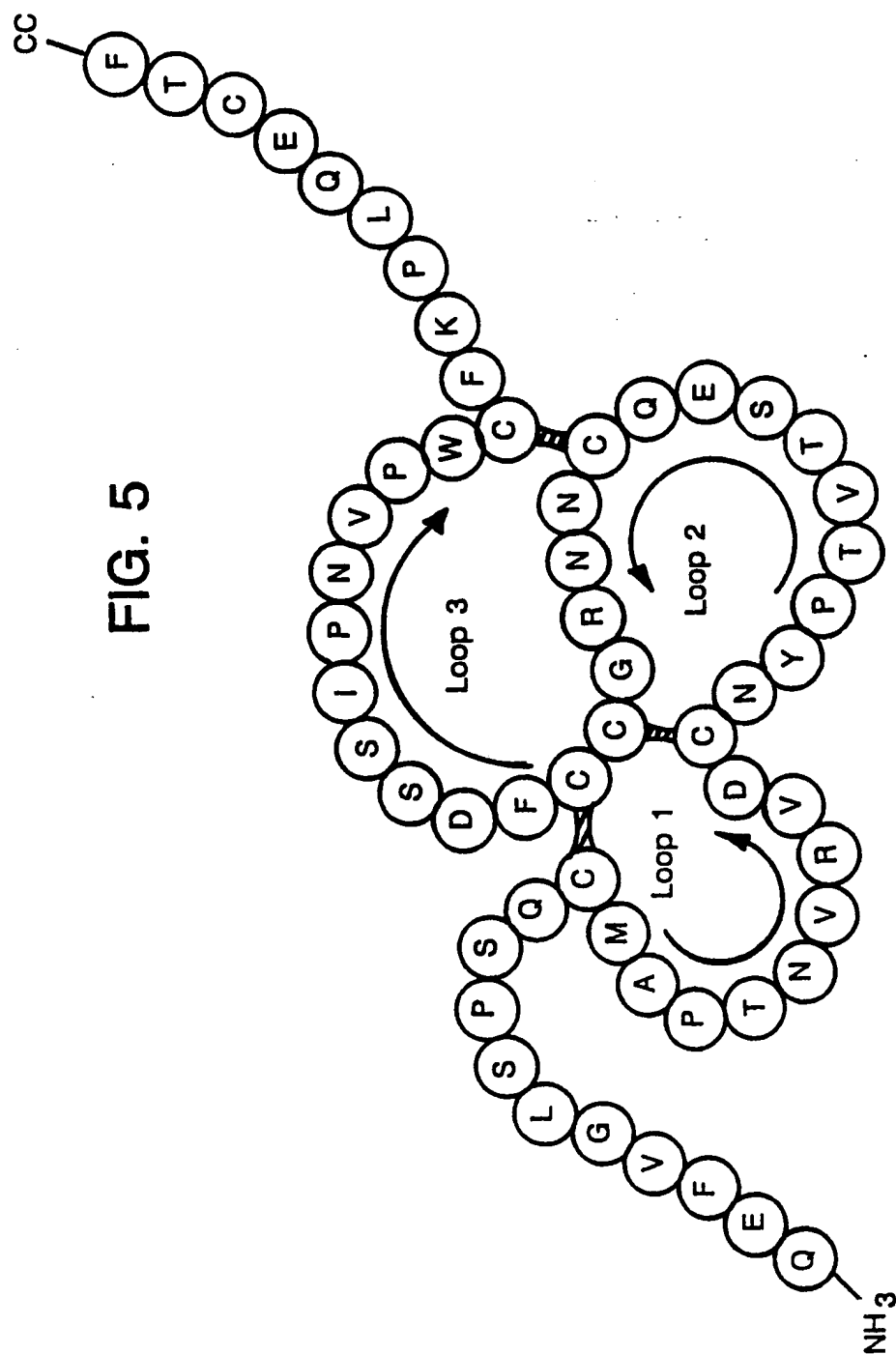


FIG. 4b





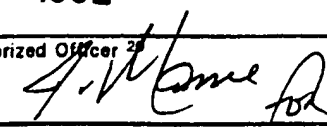
6/6

gatgctggggctggtcctggccttgctgtcctccagctctgctgaggagtacgtgggcct
-----+-----+-----+-----+-----+-----+
1 60
M L G L V L A L L S S S S A E E Y V G L
gtctgcaaaccagtggtgccgtgccggccaaggacaggggtggactgcggctacccccatgt
-----+-----+-----+-----+-----+-----+
61 120
S A N Q C A V P A K D R V D C G Y P H V
caccaccaaggagtgaacaaccggggctgctgctttgactccaggatccctggagtgcc
-----+-----+-----+-----+-----+-----+
121 180
T P K E C N N R G C C F D S R I P G V P
ttggtgtttcaagcccctgactaggaagacagaatgcaccttctgaggcacctccagctg
-----+-----+-----+-----+-----+-----+
181 240
W C F K P L T R K T R C T F *
cccctgggatgcaggctgagcacccttgcccggctgtgattgctgccaggcactgttcat
-----+-----+-----+-----+-----+-----+
241 300
ctcagtttttctgtccctttgctcccggcaagctttctgctgaaagttcatatctggagc
-----+-----+-----+-----+-----+-----+
301 360
ctgatgtcttaacgaataaagggtcccatgctccaccgAAAA
-----+-----+-----+-----+-----+-----+
361 403

FIG. 6

INTERNATIONAL SEARCH REPORT

International Application No. PCT/US92/01200

I. CLASSIFICATION OF SUBJECT MATTER (if several classification symbols apply, indicate all) ³		
According to International Patent Classification (IPC) or to both National Classification and IPC		
IPC (5): C12Q 1/00; C12N 1/21; C07K 3/00; C07H 15/12; A01N 37/18		
US CL : Please See Attached Sheet.		
II. FIELDS SEARCHED		
Minimum Documentation Searched ⁴		
Classification System	Classification Symbols	
U.S.	435/7.1, 91, 172.3, 253.3 235.1, 320.1; 530/350, 387; 536/27; 514/2;	
Documentation Searched other than Minimum Documentation to the extent that such Documents are included in the Fields Searched ⁵		
CAS Online search terms: intestinal trefoil protein, antibody, mammalian, assay		
III. DOCUMENTS CONSIDERED TO BE RELEVANT ¹⁴		
Category ¹	Citation of Document, ¹⁶ with indication, where appropriate, of the relevant passages ¹⁷	Relevant to Claim No. ¹⁸
A	Sambrook et al., "Molecular Cloning, A Laboratory Manual", 2nd Edition, published 1989 by Cold Spring Harbor Laboratory Press, (NY), pp. 8.3 to 8.52 and 12.1 to 12.29	1-8, 18-23
A	Sambrook et al., "Molecular Cloning, A Laboratory Manual", published 1989 by Cold Spring Harbor Laboratory, (NY), pp. 18.1 to 18.29	10-14, 16, 17
A	Frank Role, "Remington's Pharmaceutical Sciences", 18th Edition, published 1990 by Mack Publishing Co., (Easton, Pa.), pp. 1389-1404.	9, 15
X,P	Gastroenterology, Volume 100, No. 5, Part 2, 19-22 May 1991, S. Suemori et al., "Identification and Molecular Cloning of a New Intestinal Trefoil Growth Factor", Abstract No. A550. See the abstract.	1-8, 18-23
<div style="display: flex; justify-content: space-between;"> <div style="width: 45%;"> <p>[*] Special categories of cited documents:¹⁶</p> <p>"A" document defining the general state of the art which is not considered to be of particular relevance</p> <p>"E" earlier document but published on or after the international filing date</p> <p>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</p> <p>"O" document referring to an oral disclosure, use, exhibition or other means</p> <p>"P" document published prior to the international filing date but later than the priority date claimed</p> </div> <div style="width: 45%;"> <p>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</p> <p>"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step</p> <p>"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art</p> <p>"&" document member of the same patent family</p> </div> </div>		
IV. CERTIFICATION		
Date of the Actual Completion of the International Search ²	Date of Mailing of this International Search Report ²	
17 APRIL 1992	11 MAY 1992	
International Searching Authority ¹	Signature of Authorized Officer ²⁰	
ISA/US	Joan Ellis 	

FURTHER INFORMATION CONTINUED FROM PREVIOUS SHEETS

I. CLASSIFICATION OF SUBJECT MATTER:
US CL :

435/7.1, 91, 172.3, 253.3 235.1, 320.1; 530/350, 387; 536/27; 514/2;